

Impact of the nisin modification machinery on the transport kinetics of NisT

Marcel Lagedroste, Jens Reiners, Sander H. J. Smits & Lutz Schmitt

Article - Version of Record



Suggested Citation:

Lagedroste, M., Reiners, J., Smits, S. H. J., & Schmitt, L. (2020). Impact of the nisin modification machinery on the transport kinetics of NisT [OnlineRessource]. Scientific Reports, 10, Article 12295. <https://doi.org/10.1038/s41598-020-69225-2>

Wissen, wo das Wissen ist.



UNIVERSITÄTS- UND
LANDESBIBLIOTHEK
DÜSSELDORF

This version is available at:

URN: <https://nbn-resolving.org/urn:nbn:de:hbz:061-20251124-115210-5>

Terms of Use:

This work is licensed under the Creative Commons Attribution 4.0 International License.

For more information see: <https://creativecommons.org/licenses/by/4.0>



OPEN

Impact of the nisin modification machinery on the transport kinetics of NisT

Marcel Lagedroste¹, Jens Reiners^{1,2}, Sander H. J. Smits¹ & Lutz Schmitt^{1✉}

Lanthipeptides are ribosomally synthesized and post-translationally modified peptides containing dehydrated amino acids and (methyl-)lanthionine rings. One of the best-studied examples is nisin produced by *Lactococcus lactis*. Nisin is synthesized as a precursor peptide comprising of an N-terminal leader peptide and a C-terminal core peptide. Amongst others, the leader peptide is crucial for enzyme recognition and acts as a secretion signal for the ABC transporter NisT that secretes nisin in a proposed channeling mechanism. Here, we present an in vivo secretion analysis of this process in the presence and absence of the nisin maturation machinery, consisting of the dehydratase NisB and the cyclase NisC. Our determined apparent secretion rates of NisT show how NisB and NisC modulate the transport kinetics of NisA. Additional in vitro studies of the detergent-solubilized NisT revealed how these enzymes and the substrates again influence the activity of transporter. In summary, this study highlights the pivotal role of NisB for NisT in the secretion process.

Abbreviations

ABC transporter	ATP binding cassette transporter
CP	Core peptide
Dha	Didehydroalanine
Dhb	Didehydrobutyrine
Lan	Lanthionine
LP	Leader peptide
MeLan	Methyl-lanthionine
MS	Mass spectrometry
NBD	Nucleotide-binding domain
NisA	Precursor peptide of nisin
NisB	Dehydratase of nisin precursor peptide
NisC	Cyclase of nisin precursor peptide
NisT	Transporter of nisin precursor peptide
PTM	Post-translational modification
RP-HPLC	Reverse-phase HPLC
SEC	Size-exclusion chromatography

Many natural products (NP) produced as secondary metabolites by microorganisms can be used as pharmaceuticals (e.g. as anticancer, antibacterial or antiviral drugs)¹. One class of these NPs are ribosomally synthesized and post-translationally modified peptides (RiPPs). The RiPP family of lanthipeptides, especially those with antimicrobial activity (lantibiotics), is gaining interest as a potential alternative for antibiotics to treat harmful multidrug resistance strains such as methicillin-resistant *Staphylococcus aureus* or vancomycin-resistant *Enterococci*^{2,3}.

Lanthipeptides (LanA) are produced as precursor peptides with an N-terminal leader peptide (LP) and a C-terminal core peptide (CP)⁴. The LP serves as a signal sequence and recognition site for the modification enzymes and the export protein^{5–8}. Furthermore, the LP keeps the modified peptide (mLanA) inactive in the cytosol⁹, while the post-translational modifications (PTM) are installed within the CP and not found in the LP¹⁰.

¹Institute of Biochemistry, Heinrich Heine University Düsseldorf, Universitätsstr. 1, 40225 Düsseldorf, Germany. ²Present address: Center for Structural Studies, Heinrich Heine University Düsseldorf, Universitätsstr. 1, 40225 Düsseldorf, Germany. ✉email: lutz.schmitt@hhu.de

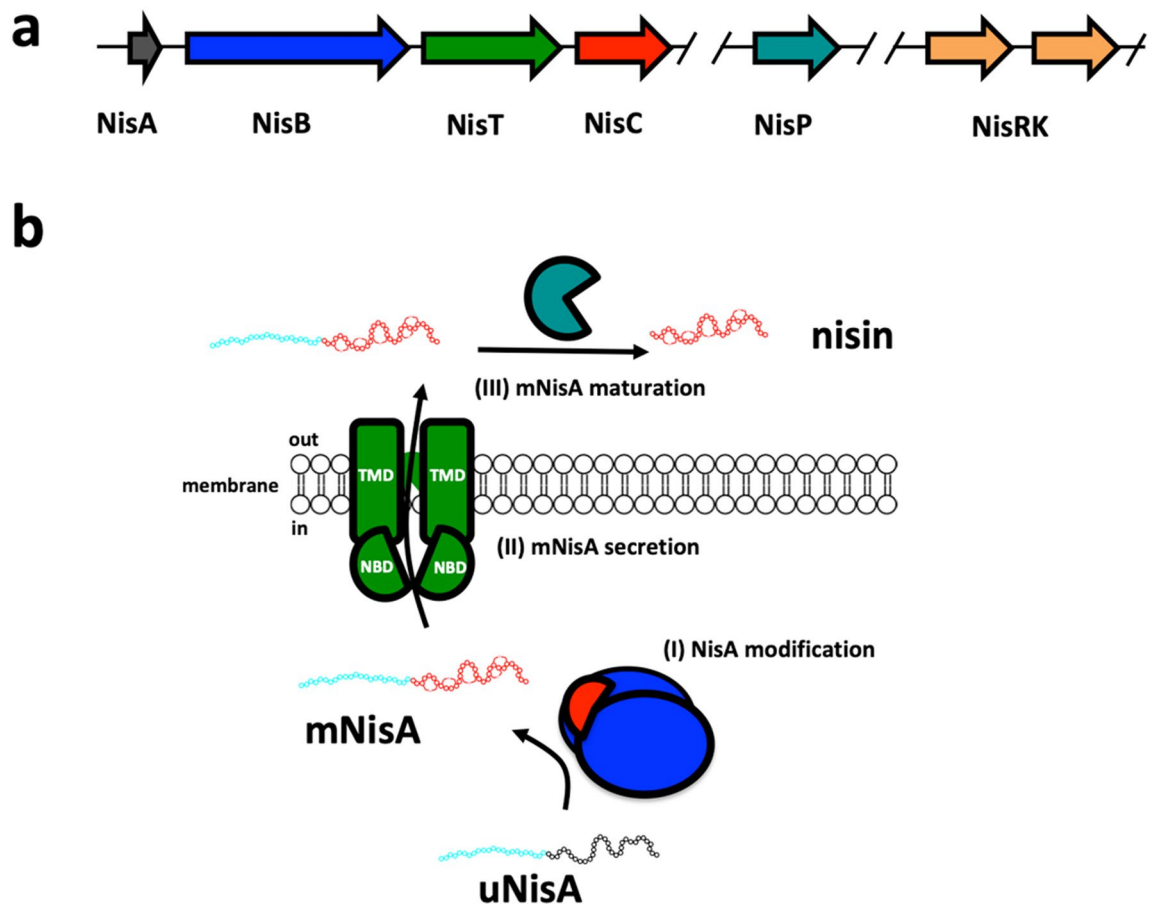


Figure 1. Scheme of nisin modification and secretion system. (a) The lanthipeptide nisin (NisA, grey) operon encodes for the modification and secretion enzymes. (b) The enzyme NisB (blue) catalyzes the dehydration reaction of unmodified NisA (uNisA), whereas NisC (red) catalyzes the thioether ring formation resulting in modified NisA (mNisA). The ABC transporter NisT (green) translocates mNisA across the membrane to the exterior. Finally, the mature peptide is processed by the serine protease NisP (turquoise) and active nisin is released. The two-component system (TCS) consisting of NisR and NisK (orange) is controlling the expression of these proteins. Please note, that the operon is partial represented and shows only proteins responsible for nisin maturation and secretion.

These PTM's are unusual amino acids such as didehydroalanine (Dha), didehydrobutyrine (Dhb) or (methyl-) lanthionine ((Me)Lan)^{11,12}.

Nisin is produced by the Gram-positive bacterium *Lactococcus lactis* as a precursor peptide (NisA), where the genes for modification, secretion and maturation enzymes are located on one operon (Fig. 1a)⁸. First, the ribosomally synthesized NisA is modified by the modification enzymes NisB and NisC (Fig. 1b, I). Within the unmodified NisA serine and threonine residues are dehydrated by the dehydratase NisB via a tRNA-dependent glutamylation and elimination reaction to Dha and Dhb residues^{13–15}. Subsequently, the dehydrated amino acid are coupled to neighboring cysteine residues via a Michael-like addition catalyzed stereo- and regio-specifically by the cyclase NisC^{16–18}. The reaction of both enzymes follows an alternating mode with a N- to C-terminus directionality yielding (Me)Lan residues^{19,20}. Next, the exporter protein NisT secretes the modified NisA (mNisA) to the exterior (Fig. 1b, II)²¹. Finally, the LP is cleaved by the extracellularly located serine protease NisP and active nisin is released (Fig. 1b, III)¹⁰.

The lanthipeptide exporters (LanT) belong to the superfamily of ABC transporters, which are found in all kingdoms of life²². Bacterial ABC transporters are composed of mainly two domains²³. One domain is the transmembrane domain (TMD) important for substrate translocation. The other domain is the nucleotide-binding domain (NBD), which binds and hydrolyses ATP to energize conformational changes used for substrate translocation. Some lanthipeptide exporters harbor an additional domain, a C39 peptidase domain. This domain is classified as a bacteriocin-processing endopeptidase²⁴ and we term this subfamily of transporters LanT_{C39P}.

All known lanthipeptide exporters function as dimers and translocation of the lanthipeptide is LP-dependent^{5,25–27}. For some LanT/ Lan_{C39P}T proteins it is proposed that the exporter and modification proteins assemble a multimeric enzyme complex at the membrane, which translocate the substrate to the exterior (e.g. nisin, subtilin and nukacin ISK-1)^{28–30}.

In 2004, it was shown that the secretion of nisin without the modification enzymes NisB, NisC and the protease NisP was possible⁵. In vivo studies on the secretion process expanded the knowledge on the nisin

modification and secretion system^{31–34}. According to these studies the observed high secretion efficiency of NisA by NisBTC was explained by a ‘channeling mechanism’^{19,35}. Other studies focused on the application of the nisin modification machinery to produce nisin variants or lantibiotics and secrete them by NisT^{36–39}. Despite the in vivo analysis of NisA secretion, a first analysis with labeled precursor peptides was performed and gave first insights into the kinetics of secretion³⁵. Nevertheless, a systematic and quantitative analysis of the secretion mechanism by determining kinetic parameters for NisA translocation by NisT is still required.

In our study, we performed an in vivo and in vitro characterization of NisT to shed light on the secretion mechanism of NisA. We determined the kinetic parameters of NisA secretion by analyzing the supernatant of NisA secreting *L. lactis* strains via reverse-phase (RP-)HPLC. The resulting apparent secretion rate (NisA·NisT^{−1}·min^{−1}) of NisT was compared with the rate of the NisBTC system and demonstrated a large enhancement in the presence of the nisin modification machinery. The in vitro characterization of NisT is the first study revealing insights into the specific activity of a LanT lanthipeptide transporter and its modification enzymes as well as its substrate. In conclusion, we could demonstrate a direct enhancement of the secretion by the maturation enzymes and a bridging function of the dehydratase NisB for the interaction of NisC and NisT.

Results

In vivo secretion assay of NisA. To obtain further insights into the mechanism of lanthipeptide secretion, the NisA secretion level of the *L. lactis* strain NZ9000 was investigated in the presence and absence of the modification machinery. Here, the well-known nisin secretion and maturation system^{19,31,35,37} was used to establish an in vivo secretion assay, where the supernatants were employed to determine the secretion level of NisA peptide via RP-HPLC analysis. In our study, we used the strain *L. lactis* NZ9000BTC (Table S1) producing fully modified NisA (mNisA) that is secreted by NisT. The secretion of other modification states of NisA can be analyzed by deletions of one of the modification enzymes or by creating inactive mutants. The mutation NisC_{H331A} (strain NZ9000BTC_{H331A}) or the deletion of NisC (strain NZ9000BT) resulted in the secretion of dehydrated NisA (dNisA). Deletion of NisB (strain NZ9000TC) or NisB and NisC (strain NZ9000T) resulted in the secretion of unmodified NisA (uNisA). The deletion of NisT (strain NZ9000BC) or the mutation of the histidine of the H-loop in the NBD of NisT (strain NZ9000BT_{H551A}C) totally abolished NisA secretion (Fig. 2). The latter two strains were defined as the background of the secretion analysis, in which NisA is expressed but not secreted. For the secretion analysis, all *L. lactis* strains were grown in minimal medium at 30 °C and after induction samples of supernatants were analyzed every hour (0–6 h). Subsequently, the supernatant was analyzed by RP-HPLC (Fig. 2, Figure S1) and the peak area of NisA peptides was determined to calculate the amount of peptide. The amount of secreted peptide was plotted against time and a non-linear fitting was applied to determine $V_{\max}$ (maximal amount of peptide; in nmol) and $K_{0.5}$ (time point of 50% secreted peptide in min). These kinetic parameters allowed a direct comparison of the secretion efficiency of the different *L. lactis* strains employed in this study.

Strain NZ9000BTC had a $V_{\max}$ value of 534 ± 44 nmol and a $K_{0.5}$ of 134 ± 12 min. It secreted NisA most efficiently in comparison to the other strains (Fig. 2, Table S3). In the cytoplasmic fraction of this strain only a small amount of mNisA was detected by Western blot at hours 2, 3 and 4 after induction (Figure S2a). This finding is similar to the previously published data for the nisin secretion and modification system³⁵.

Mutation of the H-loop at position 551 to alanine in the NBD of the ABC transporter NisT abolished the secretion of mNisA and no peptide was detected in the supernatant (Fig. 2, Figure S2c). The same result was observed for the strain NZ9000BC (Figure S2g). Here, high amounts of mNisA were detected in the cytoplasmic fraction of the strains NZ9000BT_{H551A}C and NZ9000BC (Figure S1c/g). Thus, *nisT* deletion and the H-loop mutation both abolished mNisA secretion.

Deletion of *nisC* (strain NZ9000BT) resulted in a lower secretion efficiency of dNisA and the $V_{\max}$ value of 247 ± 15 nmol and a $K_{0.5}$ of 152 ± 9 min (Fig. 2, Table S3). The amount of secreted dNisA was reduced by a factor of 2.2 in comparison to mNisA (strain NZ9000BTC). Interestingly, the mutation of the catalytic histidine residue (H331) to alanine⁴⁰ reduced the $V_{\max}$ value (168 ± 16 nmol) by factor of 3.2. The $K_{0.5}$ value increased to 200 ± 16 min (Fig. 2). The analysis of the cytoplasmic fraction of the strain NZ9000BTC_{H331A} showed a higher amount of dNisA inside the cell, which only slowly decreased over the time (Figure S2b). Slightly lower amounts of dNisA were observed in the cytoplasmic fraction of NZ9000BT (Figure S2d).

Strain NZ9000T, which was obtained after the deletion of *nisB* and *nisC*, had a slightly reduced $V_{\max}$ value of 137 ± 30 nmol with a $K_{0.5}$ of 144 ± 41 min (Fig. 2, Table S3). The secretion of uNisA was reduced by a factor of 3.9 compared to strain NZ9000BTC. The lowest amount of secreted peptide was determined in the supernatant of strain NZ9000TC with a $V_{\max}$ value of 38 ± 8 nmol. Here, a higher amount of uNisA was detected in the cytoplasmic fraction, whereas no peptide was observed in strain NZ9000T.

In all strains, NisB, NisC and NisT were detected in their corresponding fraction (cytoplasmic or membrane) (Figure S2a–g). The proteins NisB, NisC/NisC_{H331A} and NisA were observed in the cytoplasmic fractions (except uNisA from the cytoplasm of NZ9000T). NisT was detected by Western blot in the membrane fraction of all strains (Figure S2a–g). In the case of mNisA expressing strains (NZ9000BTC, NZ9000BT_{H551A}C, and NZ9000BC) all proteins were detected by Western blot even at time point zero.

Determination of the apparent secretion rate of mNisA. We measured the apparent secretion rate ($V_{s \text{ app}}$) by an in vivo secretion assay. First, the amount of secreted mNisA at different time points was determined and plotted as nmol mNisA against time (Fig. 2). Second, the amount of the ABC transporter NisT was obtained by analyzing the membrane fraction of strains NZ9000BTC and NZ9000T at each time point by Western blot (Figure S3a,b). Here, known concentrations of purified NBD protein were used as a standard to determine the amount of NisT in pmol. Combining these data, the secretion rate of NisA molecules per NisT

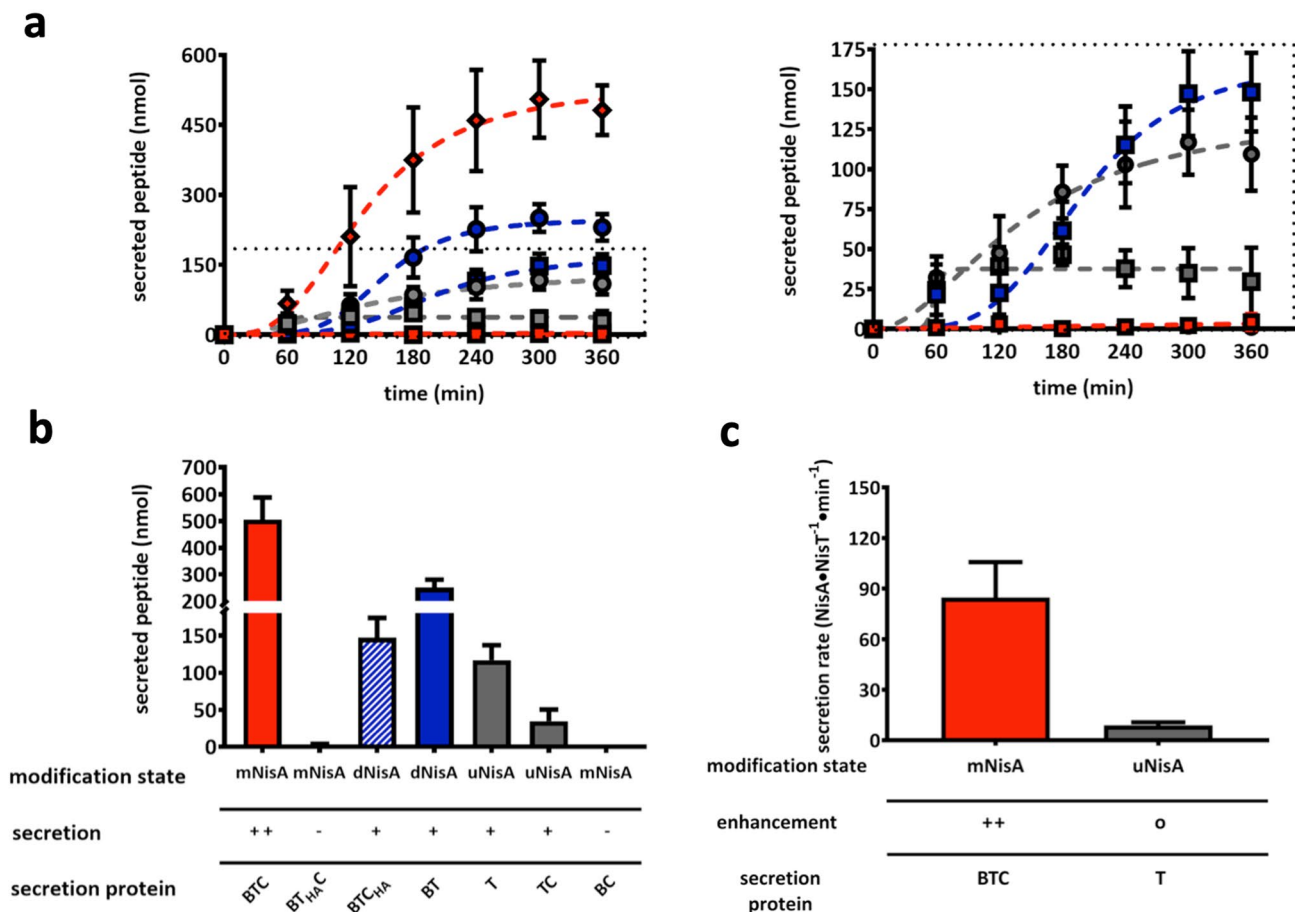


Figure 2. In vivo secretion assay of different *L. lactis* NZ9000 strains. **(a)** The supernatants of NisA secreting *L. lactis* NZ9000 strains was analyzed by RP-HPLC and the amount of NisA were determined. Amounts of secreted peptides (nmol) are plotted against time (min) and the resulting curves were fitted by an allosteric sigmoidal fit. Modified NisA (mNisA, red) was secreted by strain NZ9000BTC (red rhomb) and can be precluded by *nisT* deletion (strain NZ9000BC, clear dot) or an ATPase deficient mutant (NZ9000BT_{H551A}C, red square). Dehydrated NisA (dNisA, blue) was secreted by strains NZ9000BTC_{H331A} (blue square) and NZ9000BT (blue dots), whereas unmodified NisA (uNisA, grey) was secreted by the strains NZ9000T (grey dots) and NZ9000TC (grey square). Dashed square shows a zoom-in on strains with lower secretion level. **(b)** The kinetic parameter of $V_{\max}$ (nmol) of secreted peptides was plotted as bars against the various secretion systems. **(c)** The secretion rate of NisA molecules per NisT molecule was plotted against time (min) and fitted by linear regression. The slope represented the secretion rate of NisA·NisT⁻¹·min⁻¹ for the strains NZ9000BTC and NZ9000T. All data represent secretion experiments from at least five different transformants and are represented as means $\pm$ s.d. (n=5). ++: WT secretion; o: low secretion; -: no secretion.

molecules was calculated. The plot of nmol·NisA·NisT⁻¹ against time (min) was fitted by linear regression (Figure S3c). The slope of the linear regression corresponds to $V_{s,app}$ of NisA·NisT⁻¹·min⁻¹, which is 84.8 ± 21 for the strain NZ9000BTC (Fig. 2c, Figure S3c). A strongly reduced secretion rate was determined for the strain NZ9000T (8.8 ± 1.8) as described qualitatively previously³⁵. The determined secretion rate thus clearly demonstrated an enhancement in the presence of the modification enzymes NisB and NisC.

Purification and basal ATPase activity of NisT, a class I lanthipeptide transporter. In order to determine the in vitro activity of a lanthipeptide ABC transporter, we purified NisT the exporter of the class I lanthipeptide nisin as a deca-histidine tagged protein variant (10HNisT). 10HNisT was homologously expressed in *L. lactis* NZ9000 and purified (Fig. 3a) after solubilization with the lipid-like surfactant Fos-choline-16 (FC-16, Anatrace) by immobilized metal ion affinity (IMAC) and size-exclusion chromatography (SEC). Similar to other membrane proteins, 10HNisT (72.5 kDa) showed a higher mobility on the SDS-PAGE gel and migrated at approximately 60 kDa. 10HNisT eluted as a homogeneous peak from SEC (Fig. 3a), subsequently the main elution fractions were further concentrated to 50 μ M and used for ATPase activity assay.

For the ATPase activity assay, the detergent was exchanged to CYMAL5 (Anatrace) and the ATPase rate was expressed as specific ATPase rate (nmol·min⁻¹·mg⁻¹). The kinetic parameters of 10HNisT in detergent solution were determined and resulted in $V_{\max}$, K_m and k_{cat} values for the transporter without its substrate (basal ATPase activity). The concentration of 10HNisT was kept constant (1 μ M), whereas the concentration of ATP was varied

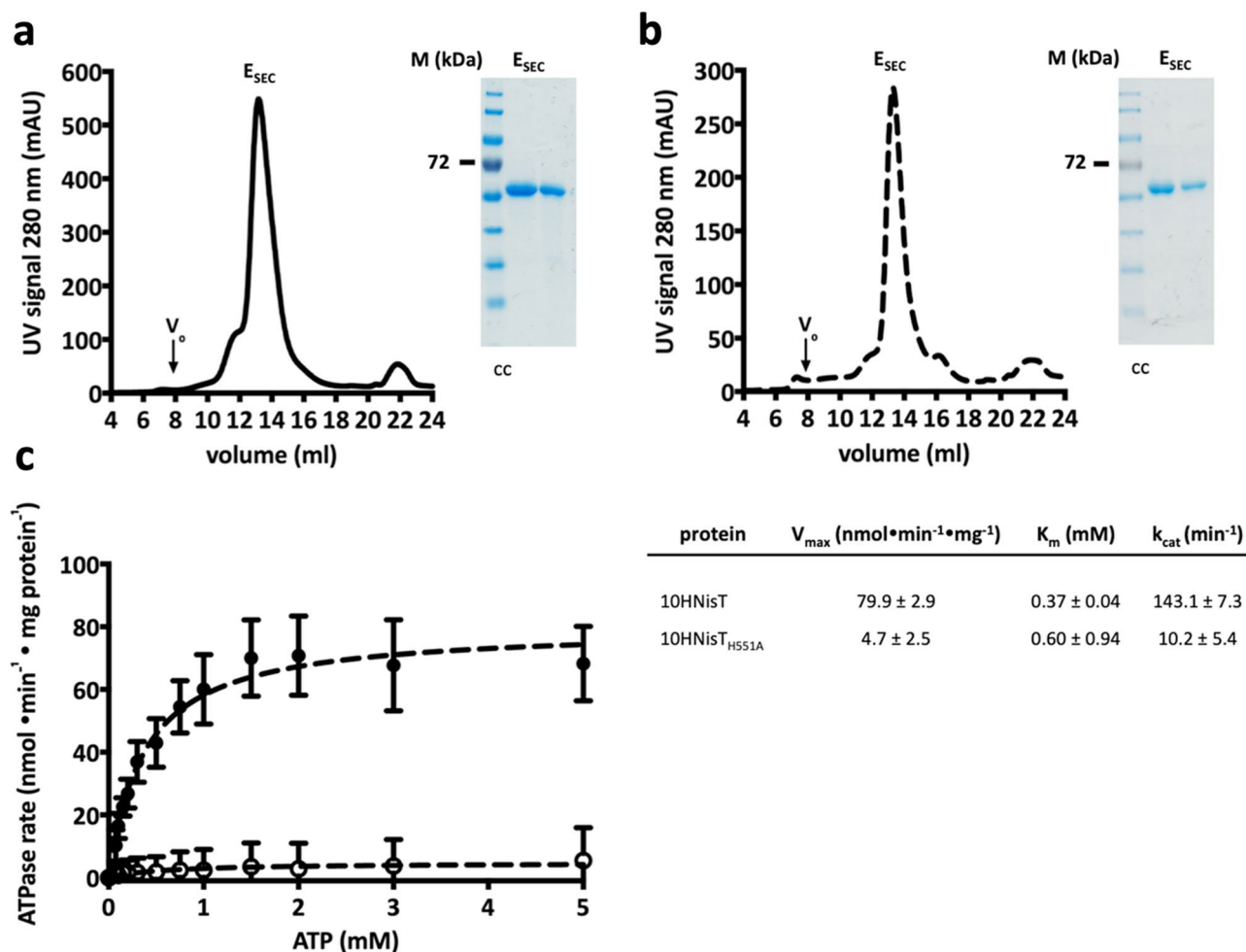


Figure 3. Purification and ATPase activity assay of NisT. **(a)** SEC chromatogram of 10HNisT (WT, black line) displayed a homogeneous peak (E_{SEC}) at 13 ml on a Superose 6 10/300 GL column (V_0 : void volume of the column). Inset: A typical colloidal Coomassie (cc) stained SDS-PAGE gel shows a protein band between 55 and 72 kDa marker protein bands (M). **(b)** Purification of the H-loop mutant 10HNisT_{H551A} (HA, dashed line) showed comparable results for SEC profile and SDS-PAGE gel (inset). **(c)** The specific ATPase rate (nmol·min⁻¹·mg protein⁻¹) of purified WT (black dot) and HA mutant (unfilled circle) was plotted against ATP concentration (mM) to determine kinetic parameters. The ATPase rate was fitted by Michaelis–Menten equation to determine $V_{\max}$ (nmol·min⁻¹·mg protein⁻¹), K_m (mM) and k_{cat} (min⁻¹). Activity assays were performed from five independent experiments with three replicates and are represented as means ± s.d. (n = 5).

from 0 to 5 mM and the reaction was stopped after 30 min. The basal ATPase rate of 10HTNisT had a $V_{\max}$ value of 79.9 ± 2.9 nmol·min⁻¹·mg⁻¹, a K_m value of 0.37 ± 0.04 mM resulting in a k_{cat} value of 143.1 ± 7.3 min⁻¹ (Fig. 3c). As a control the H-loop mutant of 10HNisT (10HNisT_{H551A}; HA-mutant) was also purified following the same protocol and used in the ATPase activity assay (Fig. 3b). The ATPase rate of the HA-mutant was reduced by a factor of 17 ($V_{\max}$ value 4.7 ± 2.5 nmol·min⁻¹·mg⁻¹). The K_m value increased by a factor of 1.62 (0.60 ± 0.94 mM), whereas the k_{cat} value was 10.2 ± 5.4 min⁻¹ (14-fold lower than WT 10HNisT) (Fig. 3c).

In vitro ATPase activity with NisA variants. To investigate the effect of substrate on the ATPase rate of 10HNisT, we added different NisA variants. First, the NisA peptides in different modification states (uNisA, dNisA and mNisA, respectively) were purified (Figure S4). Additionally, the leader peptide of NisA (NisA_{LP}) was used in the ATPase assay to evaluate whether the isolated LP is sufficient for recognition by NisT.

For the activity assay the ATP concentration was kept constant at 5 mM, while the substrate concentration was varied from 0 to 40 μ M. 10HNisT was pre-incubated with the peptides prior to the activity assay. The basal activity of 10HNisT was set to 100% and the ATPase rate with substrates was expressed as normalized ATPase rate. The ATPase rate of 10HNisT was slightly increased for all peptides. However, a concentration dependent stimulation of the transporter was not observed for all the different tested peptides (Fig. 4a,b) suggesting that the ATPase is not modulated by the NisA variants under the experimental conditions.

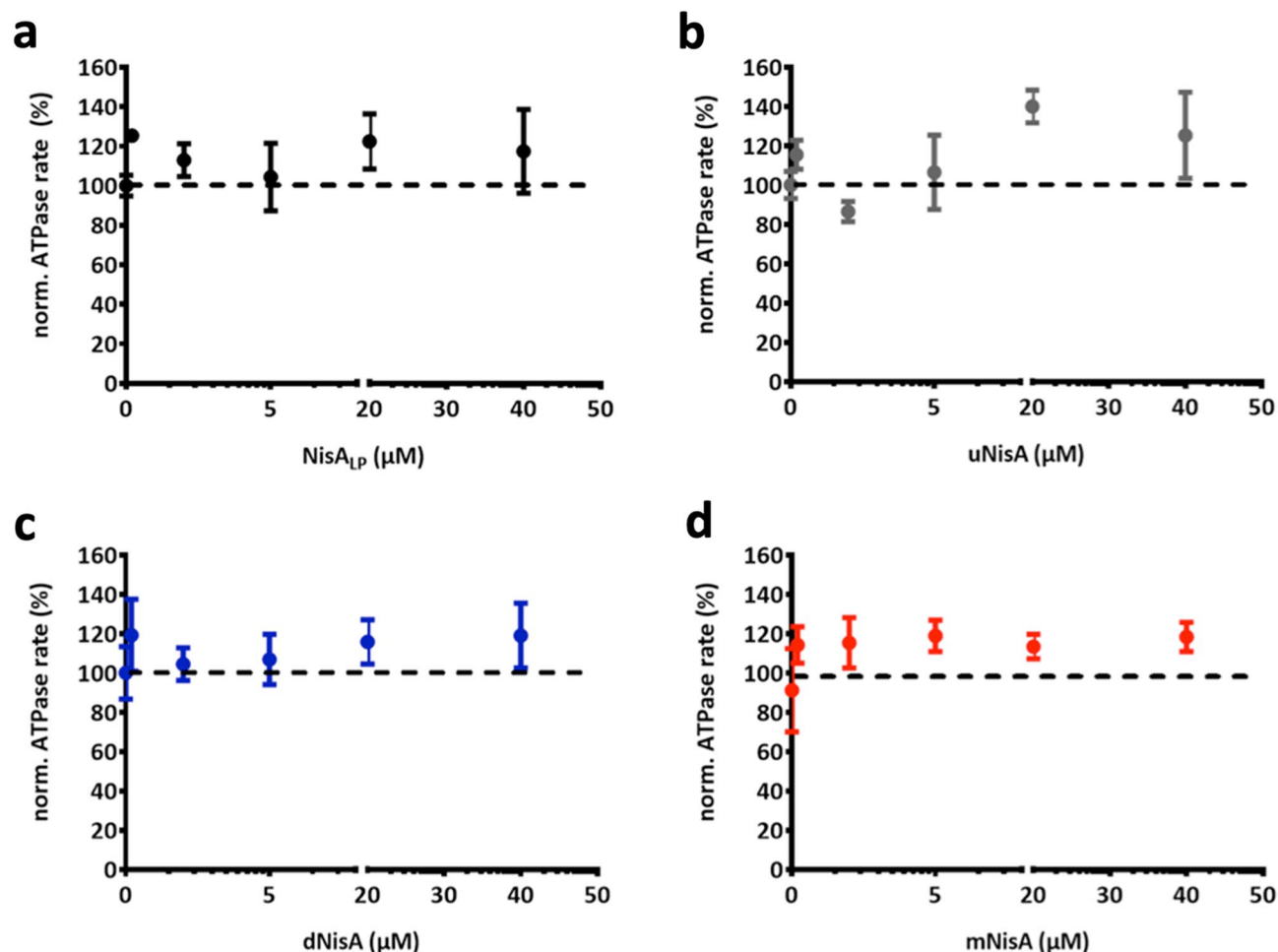


Figure 4. Dependence of NisT ATPase rate on different substrates. The ATPase rate of purified 10HNisT was analyzed in the presence of different substrates. (a) The leader peptide of NisA (NisA_{LP}, black dots), (b) uNisA (grey dots), (c) dNisA (blue dots) and (d) mNisA (red dots) was used in various concentrations (μM) and the ATPase rate is shown as normalized ATPase rate (%). The basal ATPase rate (from 10HNisT without any substrate; compare Fig. 3) was set as 100% (dashed line) and further values were normalized accordingly. The assays were performed in at least four independent experiments and are represented as means $\pm$ s.d. ($n=4$). The means were analyzed by a one-way ANOVA and the differences were not significant (p -value: ≥ 0.05). ns: not significant.

In vitro ATPase activity in presence of NisBC and mNisA. Previous studies³⁵ and our data demonstrated that the secretion of NisA is strongly enhanced by the modification proteins NisB and NisC (see in vivo secretion assay) and therefore the ATPase rate of NisT might also be influenced by these interaction partner. To investigate the effect of NisB and NisC on the ATPase rate of 10HNisT the ATPase activity assay was repeated under the same conditions (ATP concentration constant, various concentration of interaction partners). We observed that the ATPase rate of NisT was independent of the various concentrations of NisB or NisC. Thus, only fixed molar ratio of 10HNisT to the interaction partner was used (NisT:NisB/NisC 1:2; in the case of NisT:NisBC 1:2:2) (Fig. 5a). The basal ATPase rate of NisT was $62.5 \pm 9.4 \text{ nmol} \cdot \text{min}^{-1} \cdot \text{mg}^{-1}$ and was not changed within the experimental error in the presence of NisB or NisC ($54.7 \pm 4.5 \text{ nmol} \cdot \text{min}^{-1} \cdot \text{mg}^{-1}$, $59.3 \pm 4.9 \text{ nmol} \cdot \text{min}^{-1} \cdot \text{mg}^{-1}$). If both proteins were used in the assay (NisBC), the ATPase rate of 10HNisT was reduced by a factor of 1.3 ($49.3 \pm 4.4 \text{ nmol} \cdot \text{min}^{-1} \cdot \text{mg}^{-1}$), but the difference was not significant (Fig. 5a).

Next, the ATPase rate of NisT with NisBC was investigated in presence of NisA_{LP} and mNisA, at concentrations ranging from 0 to 40 μM . The ATPase rate with the substrate NisA_{LP} was slightly increased but not in a concentration dependent manner (Fig. 5b), while a decreasing effect on the ATPase rate was observed in presence of mNisA. However, a concentration dependent is not observed and the change is within the experimental error (Fig. 5c).

Interaction of NisT with NisBC. In 2017 the assembly of the nisin modification complex consisting of NisB₂C and NisA was published and shed light on the stoichiometry and structure of the complex in solution⁴¹. Additionally, an influence of the last ring on complex disassembly was determined. The next step would be

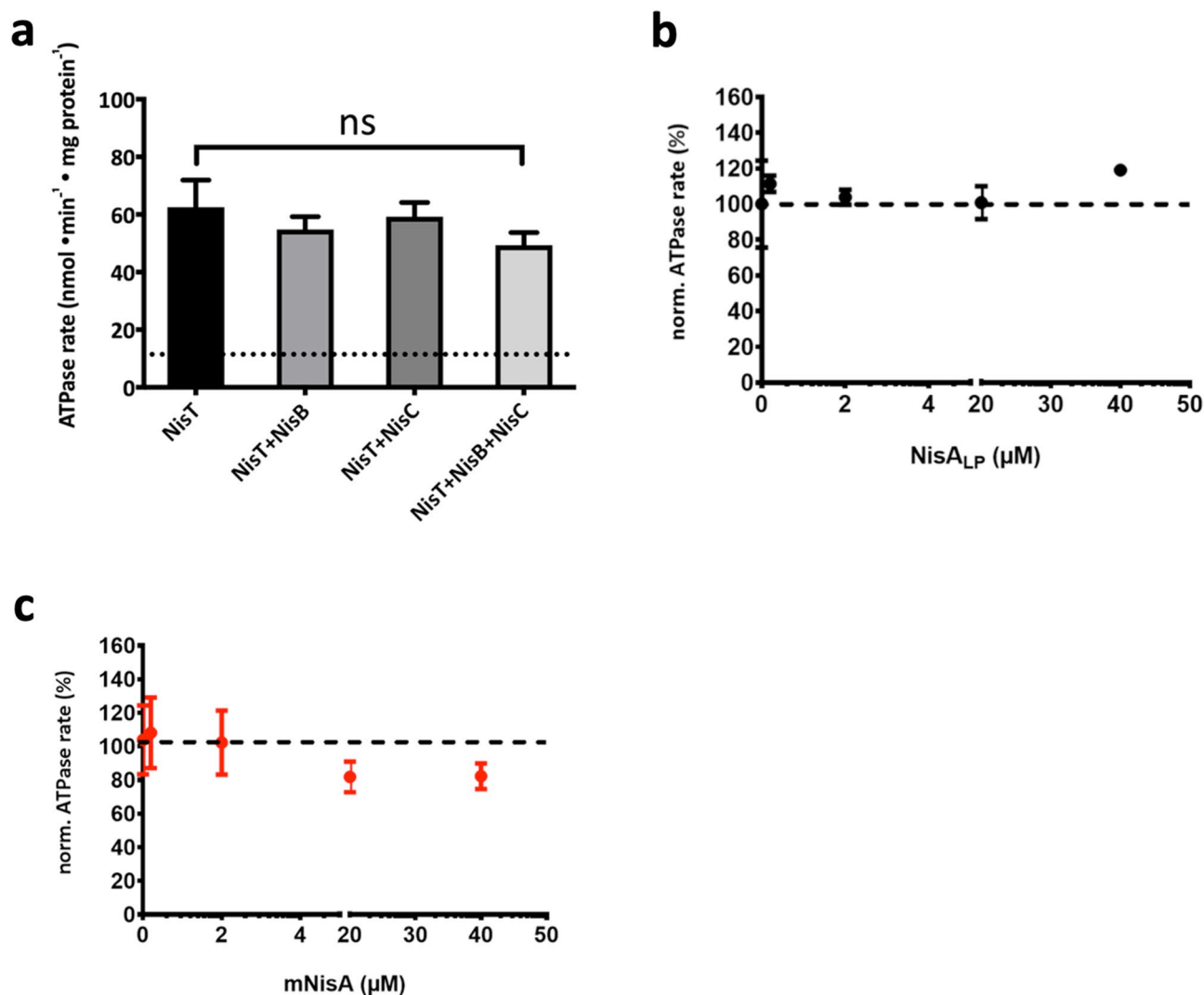


Figure 5. Influence of NisBC on the ATPase rate of NisT. The ATPase rate of purified 10HNisT was analyzed in the presence of the modification enzymes NisB and NisC, respectively. (a) ATPase rate of 10HNisT was plotted against variation of 10HNisT with the modification enzymes NisB, NisC and NisBC. It showed the normalized ATPase rate, in which the ATPase rate of 10HNisT was set to 100% (dashed line). (b) The substrate NisA_{LP} (black dots) and (c) mNisA (red dots) were used in the assay with 10HNisT in presence of NisBC. The normalized ATPase rate was plotted against various concentrations (μM), where the ATPase rate of 10HNisT plus NisBC was set to 100% (dashed line). All assay assays were performed in at least four independent experiments and are represented as means ± s.d. (n = 4). The means were analyzed by a one-way ANOVA and the differences were not significant (p-value: ≥ 0.05).

the interaction with NisT prior to secretion, as a proposed transient multimeric nisin modification/secretion complex²⁸, but detailed information about the interaction with the ABC transporter NisT is still missing.

Therefore, the interaction of NisT with NisB and NisC was investigated by a pull-down assay, in which 10HNisT was immobilized on Ni-NTA-magnetic beads (Qiagen). Initially, the interaction of NisT was tested with 10 μM NisB, NisC or NisBC. Interestingly, in all cases the interaction partner of NisT were observed in the elution fractions. This clearly shows a specific interaction of NisB and NisC with NisT independent of NisA (Fig. 6a). The controls, in which the Ni-NTA-magnetic beads were incubated with NisB and NisC without immobilized 10HNisT, revealed no protein in the elution fractions (Figure S5).

The same set up was used in the presence of the substrate mNisA_{CCCCA}, which lacks the last lanthionine-ring (ring E) and stabilizes the maturation complex of NisBC⁴¹. This substrate showed no additional effect on the interaction of NisB with NisT. However, the interaction of NisT and NisC was affected and the amount of co-eluted NisC was strongly reduced (Fig. 6b). After addition of NisB, the interaction of NisB and NisC with NisT was restored (Fig. 6b). Noteworthy, the addition of mNisA instead of the ring-mutant shows an identical result on complex formation. Unfortunately, the analysis of all elution fractions with an antibody against the LP gave no signals for the substrates mNisA/mNisA_{CCCCA}. This might be due to low concentrations of the peptides in the elution fractions.

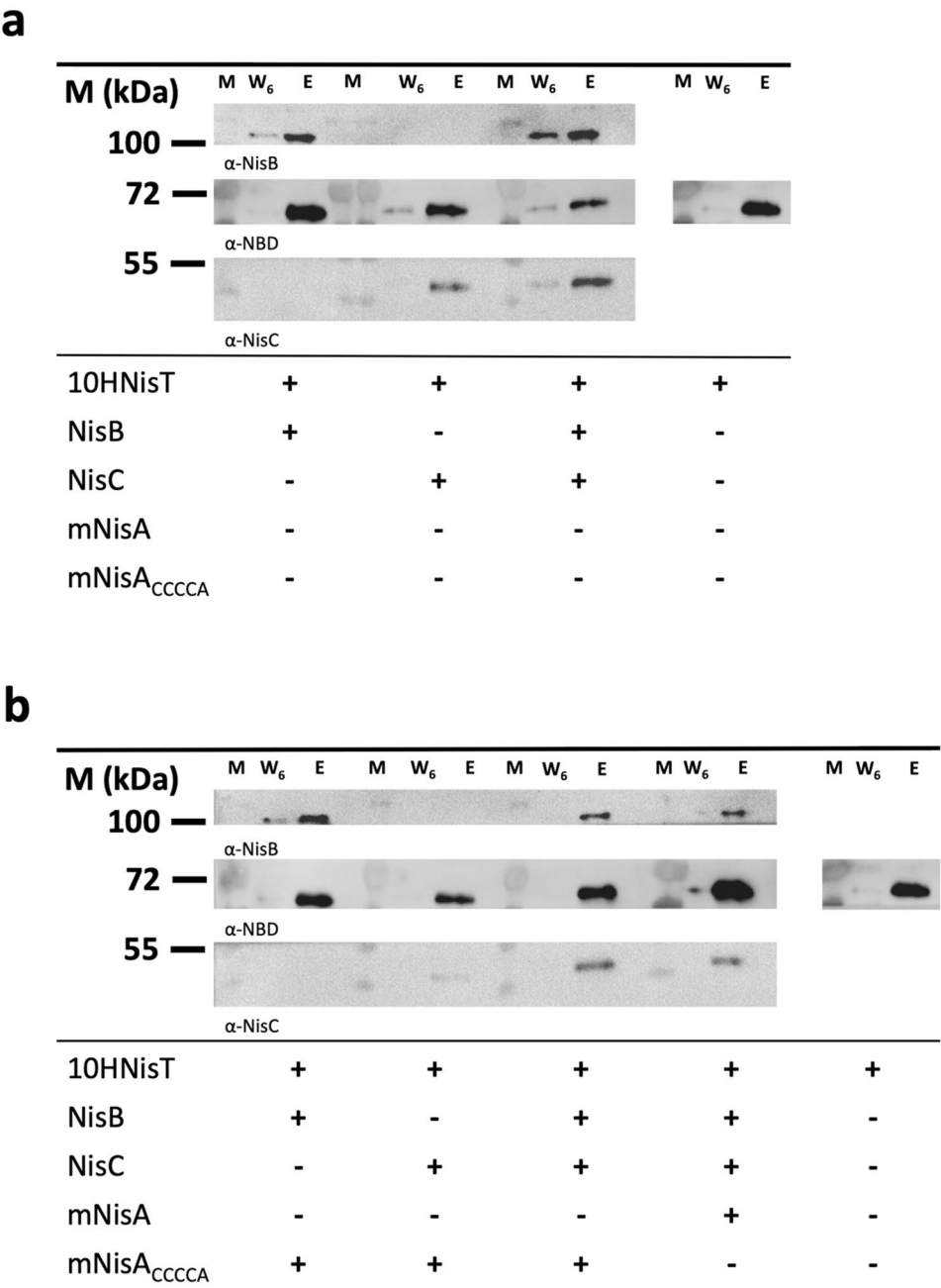


Figure 6. Pull-down assay of NisT with NisB and NisC. The interaction of 10HNisT with NisB and NisC was studied by a pull-down assay. The ABC transporter was immobilized on NTA-magnetic beads, the specific interaction partner were added and incubated. After six washing steps, the last washing step (W₆) and the EDTA elution fraction (E) were analyzed by Western blot with the specific antibodies (α-NDB, α-NisB or α-NisC). Western blots displayed the eluted bands for NisB, NisT and NisC without substrate (**a**) and with substrates mNisA or mNisA_{CCCCA} (**b**). The pull-down assay was repeated three times and showed similar results. M: marker protein bands; + : protein was used in assay; -: protein was not used in assay.

In summary, this is the first time that beside the interaction of NisT and NisC, an interaction of NisT with NisB was shown. Even the co-elution of NisB and NisC with NisT as a complex in the presence of the substrates mNisA and the ring-mutant mNisA_{CCCCA} was observed. Although no enhancement of the interaction between NisT and NisBC in presence of substrates mNisA/ mNisA_{CCCCA} was detected, NisB seems to have a pivotal role for complex formation.

Discussion

The mechanism of lanthipeptide modifications was subject to many studies, but still only little is known about the secretion process of class I lanthipeptide ABC transporters (LanT). Only a few *in vivo* studies investigated the translocation of lanthipeptides nisin, subtilin, Pep5 or epidermin^{5,25,42–44}. Amongst these systems, the nisin modification and secretion system (NisBTC) is the best studied one and is commonly employed to secrete nisin variants, lanthipeptides or non-lanthionine containing peptides^{5,31,37}. It is commonly established, that nisin is ribosomally synthesized as a precursor peptide NisA which undergoes special post-translational modifications (e.g. Dha, Dhb, Lan and MeLan)^{11,45,46}. The PTM of the CP are installed in a coordinated manner by the nisin modification complex NisB₂C^{41,47}.

Not only is the modification a tightly coupled process, but also the secretion of mNisA by NisT. A proposed channeling mechanism through interaction with NisB/NisC might explain mNisA translocation. Here, mNisA can be detected in the medium within the first minute after induction³⁵. We characterized the LanT-type transporter NisT with respect to the secretion process and its specific ATPase activity. To study the mechanism of nisin secretion, we focused on two main topics: (I) *In vivo* secretion rate of NisA by NisT and (II) *in vitro* activity of NisT with and without substrate.

In 2008 van den Berg van Saparoea et al. conducted a kinetic analysis of nisin production with the strains NZ9700 and NZ9000 transformed with a two plasmid system³⁵. They demonstrated distinct contributions of the modification enzymes NisB and NisC with respect to lanthipeptide secretion and proposed that the secretion process of NisT occurred via a channeling mechanism. This hypothesis was supported by further *in vivo* studies, in which some mechanistic aspects of NisB and NisC modification were investigated^{19,47,48}. Although these studies clearly demonstrated the dependence of NisA secretion on the modification enzymes, they did not include a determination of the underlying kinetic parameters. To determine these kinetic parameters, we quantified the amount of secreted peptide via HPLC from different time points of various NZ9000 strains.

The first two kinetic parameters were $V_{\max}$ and $K_{0.5}$, which were obtained by an allosteric sigmoidal analysis (Fig. 2). Generally, our results are consistent with the aforementioned studies, in which strain NZ9000BTC had the highest $V_{\max}$ and the lowest $K_{0.5}$ value reflecting a high secretion efficiency. The strains, which secreted uNisA (NZ9000T and NZ9000TC), show lower $V_{\max}$ and higher $K_{0.5}$ values. Interestingly, we observed in our secretion assay some aberrations with respect to dNisA secretion. The expression of a catalytic-inactive NisC (H331A mutant) in the NisBTC system (strain NZ9000BTC_{H331A}) did not restore the secretion level of dNisA to the WT level. This is in contrast to Lubelski et al., where a recovery of the NisA secretion to WT level was observed¹⁹. The secretion of dNisA by NZ9000BT has a higher $V_{\max}$ level and a lower $K_{0.5}$ value. However, one has to consider the time scale of the secretion assays, which might explain this difference. In our assay, the early kinetics of apparent secretion (also see³⁵) might pronounce the differences between the strains more clearly as an end-point determination after an overnight secretion. The precise determination of the apparent secretion efficiency revealed a descending order of the NisA secreting strains (NZ9000BTC > NZ9000BT > NZ9000BT C_{H331A} > NZ9000T > NZ9000TC). The secretion efficiency clearly shows, that mNisA is secreted at high rates by strain NZ9000BTC and every aberration of the secretion system reduced the rate at least by a factor of 2.2 (see strain NZ9000BT). The third kinetic parameter was the apparent secretion rate $V_{s,app}$ (NisA·NisT⁻¹·min⁻¹), which we determined for the strains NZ9000BTC and NZ9000NisT, in which the secretion efficiency was the highest for NisBTC system (84.8 ± 21 NisA·NisT⁻¹·min⁻¹; Fig. 2). This is 9.6-fold higher than with the transporter NisT alone (8.8 ± 1.68 NisA·NisT⁻¹·min⁻¹).

We observed similar results like other ABC transporter depended secretion system (e.g. HlyA T1SS⁴⁹) that ATP hydrolysis is essential for the secretion process. As the H551A mutant of NisT (strain NZ9000BT_{H551A}C) did not secrete mNisA. This mutant enables ATP binding and NBD dimerization but not ATP hydrolysis⁵⁰.

Thus, ATP hydrolysis is clearly important for ABC transporter mediated substrate translocation, we determined the *in vitro* activity of NisT in terms of ATPase rate without and with substrate. Here, the basal ATPase activity had a value of 79.9 ± 2.9 nmol·min⁻¹·mg⁻¹ with a K_m value of 0.37 ± 0.04 mM, which is in the range of other ABC transporters^{51–54} (Fig. 3). In comparison to NisT, the LanC_{39p}T transporter NukT has a low $V_{\max}$ value of 12.6 nmol·min⁻¹·mg⁻¹, but it is stimulated by mNukA up to max. 500% (at 50 μ M substrate). The cleaved substrate stimulates up to 200% (at 25 μ M substrate) and the unmodified substrate does not stimulate at all⁵⁵. In the case of NisT, we did not observe a substrate concentration dependent stimulation in the presence of NisA_{Lp}, uNisA, dNisA and mNisA (Fig. 4). Next, we extended the ATPase activity by addition of NisB and NisC to simulate NisBTC, in which the secretion of NisA is the most efficient (Fig. 5). The addition of Nis_{Lp} had no effect on the ATPase rate. However, the addition of mNisA revealed an inhibiting effect on the ATPase rate with increasing substrate concentration, although it was within the experimental error. Interestingly, a similar behavior was observed for PCAT1⁵². Another study on an ABC transporter homologue to PrtD from *Aquifex aeolicus* also noticed an inhibitory effect on ATPase activity after substrate addition⁵⁶. The open question is now, if NisT uses an equivalent mechanism, in which the interaction of the modification/ secretion complex inhibits the ATPase rate prior to translocation.

In 1996 a multimeric enzyme complex of NisBTC was proposed, but the isolation of such a complex was not successful²⁸. Therefore, we choose to study the specific interaction of NisT with NisB and NisC via a pull-down assay. Such a pull-down assay was performed with His-tagged NisA, where NisB and NisC were co-eluted from cytoplasmic fraction^{47,57}. In our study, we expanded this set up and used purified NisT, NisB, NisC and NisA (Fig. 6). We observed a specific interaction of NisT with NisC, which is in line with previously observed interaction of NisT with NisC via co-immunoprecipitation and yeast two-hybrid assay²⁸. Another specific interaction of the modification enzyme with the ABC transporter was shown for NukM and NukT. Here, the C-terminal domain of LanM (LanC-like domain; amino acids 480–917) interacts with TMD and NBD of NukT, but not with the C39 peptidase domain³⁰. Besides the interaction of NisT with NisC, we also noticed an interaction

of NisT with NisB, which was not observed in the above-mentioned study, but for SpaT and SpaB in a similar experiment²⁹. Since secretion of dNisA by NZ9000BT was observed⁵, an interaction of NisT and NisB was also proposed. Furthermore, we observed for the first time the co-elution of NisBC with NisT in the proposed lanthionine synthetase complex. Remarkably, the co-elution of transporter with the modification enzymes is not increased by addition of the substrates mNisA or mNisA_{CCCCA}. Similar amounts of the enzymes were co-eluted and we conclude that the interaction of NisT with NisB and NisC is independent of substrate. One exception was the addition of mNisA_{CCCCA} to NisT/NisC, in which the amount of co-elute NisC was reduced. Only the addition of NisB to the sample increased the co-elution of NisC. It is now commonly accepted that NisB represents the main component of the NisBTC modification/secretion complex^{19,35} and based on our data, NisB stabilizes the NisBTC complex.

In summary, we have determined the kinetic parameters for the *in vivo* secretion of nisin peptides by the modification and secretion complex NisBTC. We demonstrated that alterations in the NisBTC system lead to impaired secretion of NisA or even to no secretion, when an ATP hydrolysis deficient mutant of NisT was used. For an efficient secretion by NisT the modification enzymes NisB and NisC are prerequisite and their interaction with NisT enhances the secretion process directly by the proposed channeling mechanism³⁵. Interestingly, it is the function not the mere presence of NisC that is required, as the secretion of NisBTC with the catalytically inactive mutant of NisC did not restore the secretion. Additionally, the *in vitro* activity data of NisT demonstrate that the lanthipeptide exporter is not stimulated by the interaction partner and substrate, respectively. The observed complex formation of NisB and NisC with NisT hint to the proposed lanthionine synthetase complex, in which NisT undergoes a transition to a transport competent exporter in presence of only NisB, NisC and the substrate mNisA.

Methods

Bacterial strains and growth conditions. Strains of *Escherichia coli* and *Lactococcus lactis* and plasmids used in this study are listed in Table S1. The strains *E. coli* DH5α or BL21 were grown in LB medium at 37 °C under aerobic conditions with appropriate antibiotics (30 µg/ml kanamycin or 100 µg/ml ampicillin). The transformation of *E. coli* strains was performed following standard procedures.

The strain *L. lactis* NZ9000 (and its variants) was grown in M17⁵⁸ or minimal medium (MM)^{31,59} at 30 °C under semi aerobic conditions supplemented with 0.5% glucose (GM17/ GMM) and appropriate antibiotics (erythromycin or/and chloramphenicol at a final concentration of 5 µg/ml). To MM a vitamin mix (100 × stock solution, 1 × final) was added³¹.

For transformation of *L. lactis* NZ9000 with the expression plasmids a standard procedure for preparation of competent cells and electroporation was used⁶⁰.

Cloning of nisT and nisT variants. A nucleotide sequence for a MCS with 10H nucleotide sequence (5'–3': CAAAATAAATTATAAGGAGGCACTCAATGCATCATCATCATCATCATCATCATCATGATTATGATATTCGACCACCGAAAACCTGTATTTTCAGGGCCCATGGCATATGCATATGTCTAGACACCACCACCACCACCTGAGATCCG) was ordered as a codon-optimized, synthetic gene fragment from Life Technologies to insert it into the pNZ-SV plasmid⁶¹. The synthetic gene fragment were amplified by Phusion DNA polymerase (NEB) with the primer pair 10Hfor and 10Hrev (Table S4) for Gibson assembly. The plasmid pNZ-SV was amplified by Phusion DNA polymerase (NEB) with the primer pair infupNZ-SVfor and infupNZ-SVrev (Table S4) to linearize the vector. The gene fragment and the vector pNZ-SV were employed in the Gibson assembly by following the manufactures instructions (NEB). The Gibson assembly reactions were transformed into *E. coli* DH5α. The sequence of the construct pNZ-SV10H (Table S5) was verified by DNA sequencing (Microsynth Seqlab).

The *nisT* gene (accession number: Q03203) was amplified using genomic DNA from *L. lactis* NZ97000⁸ as a template. Phusion DNA polymerase (NEB) with the primer pair infunisTfor and infunisTrev (Table S4) was used to create overhang sequences for Gibson assembly. The plasmid pNZ-SV10H was amplified by Phusion DNA polymerase (NEB) with the primer pair linpNZ-SVfor and linpNZ-SVrev (Table S4) to linearize the vector. Subsequently, the gene and the linearized vector pNZ-SV10H were employed in the Gibson assembly and the reactions were transformed into *E. coli* DH5α. The sequence of the construct pNZ-SV10HnisT (Table S5) was verified by DNA sequencing (Microsynth Seqlab).

To generate the plasmid pIL-SVnisT the *nisT* gene from pNZ-SV10HnisT was amplified by Phusion DNA polymerase (NEB) with the primer pair infupIL-SVfor and infupIL-SVrev (Table S4) to create overhanging sequences. The plasmid pIL-SV⁶² was linearized by Phusion DNA polymerase (NEB) with the primer pair term-pNZfor and pnisArev (Table S4). The gene with overhang sequences and the vector was employed in Gibson assembly. The Gibson assembly reactions were transformed into *E. coli* DH5α. Additionally, the 10H nucleotide sequence was deleted by Phusion DNA polymerase (NEB) with the primer pair 10Hfor and infupNZ-SVrev (Table S4). The sequence of the construct pIL-SVnisT (Table S5) was verified by DNA sequencing (Microsynth Seqlab).

To generate the *nisT*_{H551A} mutant, a polymerase chain reaction using Pfu DNA polymerase (Thermo Fischer Scientific) or Pfu DNA polymerase (Promega), the template pNZ-SV10HnisT or pIL-SVnisT and the primer pair nisT_{H551A}for and nisT_{H551A}rev (Table S4) was performed according to standard procedures. The sequence of the constructs (Table S5) was verified by DNA sequencing (Microsynth Seqlab).

The plasmid pNZ-SVnisTNBD_{H348} was obtained by the deletion of the TMD sequence (1–347) from the plasmid pNV-SV10HnisT. The plasmid was amplified with Phusion DNA polymerase (NEB) with the primer pair ΔnisT_{TMD}for and ΔnisT_{TMD}rev (Table S4). The linear vector was ligated with T4-ligase (NEB) and transformed

into *E. coli* DH5α. The sequence of the construct pNZ-SVnisTNBD_{H348} (Table S5) was verified by DNA sequencing (Microsynth Seqlab).

Cloning of nisBTC and nisBTC variants. The plasmid pIL-SVnisBTC was generated from pIL-SV and pIL3BTC³¹. The plasmid pIL3BTC was digested with the restriction enzymes NotI (NEB) and BstXI (NEB) to receive a fragment BTC containing the genes *nisB*, *nisT* and *nisC*. Next, pIL-SV⁶² was also digested with NotI and BstXI (pIL-SV**). The fragment BTC and pIL-SV** were ligated with T4-ligase (NEB) and transformed into *E. coli* DH5α. The sequence of the construct pIL-SVnisBTC (Table S5) was verified by DNA sequencing (Microsynth Seqlab).

By using Phusion DNA polymerase (NEB) with the appropriate primer pairs (Table S4) the gene deletions of *nisB*, *nisC* or *nisT* were performed to generate pIL-SVnisBTC derivatives. Subsequently, the linearized vectors were ligated with T4-ligase (NEB) and transformed into *E. coli* DH5α. The sequence of the constructs (Table S5) were verified by DNA sequencing (Microsynth Seqlab).

To generate *nisT*_{H551A} and *nisC*_{H331A} mutants, a polymerase chain reaction using PfuUltra II Fusion DNA polymerase (Agilent Technologies), the template pIL-SVnisBTC and the appropriate pair of oligonucleotides (Table S4) was performed according to standard procedures. The sequence of the new constructs (Table S5) were verified by DNA sequencing (Microsynth Seqlab).

In vivo secretion assay: expression and secretion of NisA. Strain *L. lactis* NZ9000 harboring the plasmids pIL-SVnisBTC and pNZ-SVnisA hereupon termed NZ9000BTC (Table S1) was used to investigate the in vivo secretion activity of the nisin modification and secretion system (NisBTC). The use of pIL-SVnisBTC derivatives and pNZ-SVnisA for transformation into *L. lactis* NZ9000 led to strains described in detail in Table S1. For each secretion experiment new transformants were prepared and used to inoculate GM17 (Erm + Cm) with one colony. The overnight culture was centrifuged at 4,000×g for 20 min and cells were resuspended in GMM. Subsequently, 0.5 l GMM (Erm + Cm) were inoculated to OD₆₀₀ of 0.3 and incubated at 30 °C. After 60–90 min the culture (OD₆₀₀ of 0.4–0.5) was induced with 10 ng/ml nisin (powder from Sigma-Aldrich dissolved in 50 mM lactic acid). A 50 ml sample before induction (0 h) and every other hour (1–6 h) was taken. For each sample the cell were harvested by centrifugation at 4,000×g for 20 min. Subsequently, the cells were resuspended in R-buffer (50 mM Na-Phosphate buffer, pH 8, 100 mM KCl, 20% glycerol) to an OD₆₀₀ of 200, flash frozen in liquid nitrogen (N₂) and stored at –80 °C until further use. The supernatant was additionally centrifuged at 17,000×g for 20 min at 8 °C. Supernatants were kept on ice before the RP-HPLC analysis. Furthermore, 2 ml or 10 ml of the supernatant were precipitated by 1/10 volume (10%) TCA. TCA samples were incubated at 8 °C overnight. The TCA-precipitated peptide was centrifuged at 17,000×g for 20 min at 8 °C and consecutively washed three-times with ice-cold acetone. The pellets were vacuum-dried and resuspended in 60 µl per OD₆₀₀ 1 of 1×SDS-PAGE loading dye containing 5 mM β-mercaptoethanol (β-ME). These resuspended TCA-pellets were analyzed by Tricine-SDS-PAGE and Western blot.

In vivo secretion assay: analysis of cell pellets. The resuspended cell pellets were thawed on ice and 1/3 (w/v) glass beads (0.3 mm diameter) were added. Cells were disrupted on a vortex-shaker (Disruptor Genie, Scientific Industries). A cycle of 2 min disruption and 1 min incubation on ice was repeated five times. A low spin step at 17,000×g for 30 min at 8 °C and subsequently a high spin step at 100,000×g for 120 min at 8 °C was performed. The supernatant of the latter centrifugation step represents the cytoplasmic fraction and the pellet corresponds to the membrane fraction. The SDS-PAGE samples of cytoplasmic and membrane fractions were prepared by adding 4× SDS-PAGE loading dye containing 5 mM β-ME and used for SDS-PAGE as well as Western blot analysis.

In vivo secretion assay: analysis of culture supernatant. The culture supernatants containing NisA variants were analyzed by RP-HPLC (Agilent Technologies 1260 Infinity II). A LiChrospher WP 300 RP-18 end-capped column and an acetonitrile/water solvent system were used as described previously⁷. In the case of modified NisA 300 µl and for all other variants (dehydrated/unmodified) 500 µl sample were injected. Prior to the gradient (20–50% acetonitrile) a washing step of 20% acetonitrile was used to remove most of the casein peptides. The peptide amount or NisA in the supernatant was determined using the peak area integration analyzed with the Agilent Lab Advisor software. In the case of the unmodified peptide (uNisA) a retention time from 29 to 33 min, for dehydrated peptide (dNisA) from 31 to 38 min and for modified peptide (mNisA) from 33 to 38 min was used for peak integration peak. A calibration with known amounts of nisin or insulin chain B was used to obtain a linear regression line. Unknown amounts in the in vivo secretion assay samples were calculated as nmol or µM based on this linear regression line.

In vivo secretion assay: determination of kinetic parameter. The amount of secreted NisA of the different *L. lactis* NZ9000 strains were plotted against time and fitted using an allosteric sigmoidal fit (1). Note that y is the amount of secreted peptide (nmol), V_{max} the maximal secreted amount, x is time (min), K_{0.5} the time point at which 50% of V_{max} is present and h is the Hill slope indicating cooperativity. The analysis was performed using Prism 7.0c (GraphPad).

$$y = V_{\max} \frac{X^h}{K_{0.5}^h + X^h} \quad (1)$$

The apparent secretion rate ($V_{s,app}$) was determined by plotting the amount of NisA and NisT against time (min). The values were fitted using a linear regression (2). Note that y is the amount of NisA molecules per NisT molecules ($\text{NisA} \cdot \text{NisT}^{-1}$), m is the slope $V_{s,app}$ ($\text{NisA} \cdot \text{NisT}^{-1} \cdot \text{min}^{-1}$), x is the time (min), and b the y -axis interception. The analysis was performed using Prism 7.0c (GraphPad).

$$y = mx + b \quad (2)$$

Expression and purification of NisT. *L. lactis* NZ9000 strain was transformed with pNZ-SV10HnisT and placed on SGM17 (0.5 M sucrose, 0.5% glucose, 22 g/l agarose in M17 medium) agar plates containing 5 μg /ml erythromycin. A GM17 (Erm) overnight culture was inoculated with one colony and incubated at 30 °C. A GM17 (Erm) main culture was inoculated to an OD_{600} of 0.1 with the overnight culture. After 3 h incubation, expression was induced by adding 10 ng/ml nisin (powder from Sigma-Aldrich dissolved in 50 mM lactic acid) and further grown for additional 3 h. Cells were harvested by centrifugation at $4,000 \times g$ for 20 min at 8 °C and resuspended in R-buffer (50 mM Na-Phosphate buffer, pH 8, 100 mM KCl, 20% glycerol) to an OD_{600} of 200. To the resuspended cells 10 mg/ml lysozyme was added and incubated at 30 °C for 30 min. Prior to cell disruption, cells were incubated on ice for 15 min. The cell suspension was passed through a homogenizer (M-110B, Microfluidics System) at 1.5 kbar at least four times. The homogenized cell suspension was centrifuged at $12,000 \times g$ for 30 min at 8 °C. Subsequently, the supernatant was centrifuged at $100,000 \times g$ for 120 min at 4 °C to collect the membrane fraction. Membranes were resuspended with R-buffer containing 10 mM imidazole and 0.5 mM AEBSF. The total membrane protein concentration was measured by BCA assay (Thermo Fischer Scientific) and the concentration was adjusted to 5–7.5 mg/ml. Membranes were solubilized with 1% (w/v) of the lipid-like detergents FC-16 (Anatrace) for 1 h at 8 °C. Insoluble material was removed by centrifugation at $100,000 \times g$ for 30 min at 4 °C. The supernatant was applied to a 5 ml IMAC (Immobilized Metal-Ion-Affinity Chromatography) HiTrap Chelating column (GE Healthcare) preloaded with 100 mM zinc sulphate and equilibrated with low IMAC1 buffer (50 mM Na-Phosphate buffer, pH 8, 100 mM KCl, 20% glycerol and 10 mM imidazole) containing 0.5 mM AEBSF and 0.005% FC-16. Consecutively, non-bound protein was washed and the buffer was exchanged to low IMAC2 buffer (50 mM Tris-HCl pH 8, 100 mM KCl, 10% glycerol, 10 mM imidazole, 0.5 mM AEBSF and 0.005% FC-16). After an additional washing step with 30% high IMAC buffer (50 mM Tris-HCl pH 8, 100 mM KCl, 10% glycerol, 150 mM histidine, 0.5 mM AEBSF and 0.005% FC-16), 10HNisT was eluted with 100% high IMAC buffer. The elution fractions containing NisT were pooled, 10 mM DTT was added and further concentrated with a Vivaspin20 100 kDa molecular weight cut off (MWCO) centrifugal concentrator (Sartorius AG). Next, a size exclusion chromatography (SEC) was performed, where the concentrated protein sample was applied onto a Superose 6 10/300 GL column (GE Healthcare) equilibrated with SEC buffer (25 mM Tris-HCl pH 8, 50 mM KCl, 10% glycerol, 0.5 mM AEBSF, 2 mM DTT and 0.0015% FC-16). The main peak fractions were analyzed via SDS-PAGE and further concentrated via a Vivaspin6 100 kDa MWCO centrifugal concentrator (Sartorius AG) until a concentration of 50 μM was reached. The protein concentration was determined by NanoDrop spectrophotometer (Thermo Fischer Scientific) using a molar extinction coefficient of $86,180 \text{ M}^{-1} \text{ cm}^{-1}$ and the molecular mass of 72.6 kDa. Aliquots of 50 μM 10HNisT were flash frozen in liquid N_2 and stored at -80 °C until further use. The NisT variants 10HNisT_{H551A} was expressed following the same protocol.

In vitro ATPase activity assay. The ATPase activity of NisT was determined with the malachite green assay as described previously with experimental alterations⁶³. In this assay the release of inorganic orthophosphate after ATP hydrolysis was colorimetrically quantified based on a Na_2HPO_4 standard curve.

All reactions were performed at 30 °C in a total volume of 30 μl in activity assay buffer containing 0.4% CYMAL5 and 10 mM MgCl_2 .

In each reaction $\sim 2 \mu\text{g}$ of detergent-solubilized and purified NisT was used and the reaction was started by adding ATP (0–5 mM). The background of the reaction was a sample without MgCl_2 . After 30 min the reaction was stopped by transferring 25 μl of each reaction into a 96-well plate containing 175 μl stop-solution (20 mM sulphuric acid). Consecutively, 50 μl of a staining solution (0.096% (w/v) malachite green, 1.48% (w/v) ammonium heptamolybdate and 0.173% (w/v) Tween-20 in 2.36 M sulphuric acid) was added. After 10 min the amount of free inorganic orthophosphate was quantified by measuring the absorption at 595 nm using an iMark microplate reader (Bio-Rad).

The specific ATPase activity of NisT was plotted against ATP concentrations and fitted using the Michaelis–Menten Eq. (3). Note that y is the reaction velocity, $V_{\max}$ the maximal reaction velocity, x is the substrate concentration and K_m the Michaelis–Menten constant. The analysis was performed using Prism 7.0c (GraphPad).

$$y = V_{\max} \frac{X}{K_m + X} \quad (3)$$

For the reactions with substrates (NisA variants) or interaction partner (NisB and NisC) NisT was pre-incubated at 30 °C for 10 min before ATP was added to start the reaction. All reactions were performed at 30 °C in a total volume of 30 μl in activity assay buffer containing 0.4% CYMAL5, 400 mM glutamate and 10 mM MgCl_2 . In each reaction $\sim 2 \mu\text{g}$ of detergent-solubilized and purified NisT was used and the reaction was started by adding 5 mM ATP and stopped after 15 min following the procedure described above. In this reaction the concentration of the different substrates (0–40 μM) and/or interaction partner was varied and the ATPase activity was normalized to the specific ATPase activity of NisT without substrate/interaction partner. In these cases, the background was subtracted prior to normalization.

In vitro pull-down assay. The immobilization of 10HNisT to Ni-NTA magnetic beads (Quiagen) was performed as described in the manufacturer's manual. In brief, ~15 µg 10HNisT was incubated with Ni-NTA magnetic beads for 30 min at 30 °C. Excess of protein was removed by three washing steps with activity assay buffer containing 0.4% CYMAL5 and 400 mM glutamate. 10 µM of the interaction partners NisC and NisB were incubated in 1:1 molar ratio (but >10 × molar excess to NisT) separately with or without mNisA/ mNisA_{CCCCA} (20 × molar excess to NisT) in activity assay buffer containing 0.4% CYMAL5, 400 mM glutamate and 5 mM MgATP for 15 min on ice. Next, interaction partner were added to 10HNisT immobilized to Ni-NTA magnetic beads and incubated for 1 h at 30 °C. Positive control (only 10HNisT) and negative control (NisB, NisC samples were prepared by incubating the proteins with Ni-NTA magnetic beads separately. After binding the Ni-NTA magnetic beads were washed six times with activity assay buffer. Finally, 10HNisT was eluted by adding activity assay buffer containing 50 mM EDTA. The SDS-PAGE samples of pull-down assay fractions were prepared by adding 4 × SDS-PAGE loading dye containing 5 mM β-ME and used for Western blot analysis.

Received: 26 March 2020; Accepted: 6 July 2020

Published online: 23 July 2020

References

- Newman, D. J. & Cragg, G. M. Natural products as sources of new drugs from 1981 to 2014. *J. Nat. Prod.* **79**, 629–661. <https://doi.org/10.1021/acs.jnatprod.5b01055> (2016).
- Hudson, G. A. & Mitchell, D. A. RiPP antibiotics: biosynthesis and engineering potential. *Curr. Opin. Microbiol.* **45**, 61–69. <https://doi.org/10.1016/j.mib.2018.02.010> (2018).
- Dischinger, J., Basi Chipalu, S. & Bierbaum, G. Lantibiotics: promising candidates for future applications in health care. *Int. J. Med. Microbiol.* **304**, 51–62. <https://doi.org/10.1016/j.ijmm.2013.09.003> (2014).
- Arnison, P. G. *et al.* Ribosomally synthesized and post-translationally modified peptide natural products: overview and recommendations for a universal nomenclature. *Nat. Prod. Rep.* **30**, 108–160. <https://doi.org/10.1039/c2np20085f> (2013).
- Kuipers, O. P. *et al.* NisT, the transporter of the lantibiotic nisin, can transport fully modified, dehydrated, and unmodified prenisin and fusions of the leader peptide with non-lantibiotic peptides. *J. Biol. Chem.* **279**, 22176–22182. <https://doi.org/10.1074/jbc.M312789200> (2004).
- Mavaro, A. *et al.* Substrate recognition and specificity of the NisB protein, the lantibiotic dehydratase involved in nisin biosynthesis. *J. Biol. Chem.* **286**, 30552–30560. <https://doi.org/10.1074/jbc.M111.263210> (2011).
- Abts, A., Montalban-Lopez, M., Kuipers, O. P., Smits, S. H. & Schmitt, L. NisC binds the FxLx motif of the nisin leader peptide. *Biochemistry* **52**, 5387–5395. <https://doi.org/10.1021/bi4008116> (2013).
- Kuipers, O. P., Beerthuyzen, M. M., Siezen, R. J. & De Vos, W. M. Characterization of the nisin gene cluster nisABTCIPR of *Lactococcus lactis*. Requirement of expression of the nisA and nisI genes for development of immunity. *Eur. J. Biochem.* **216**, 281–291 (1993).
- Van der Meer, J. R. *et al.* Influence of amino acid substitutions in the nisin leader peptide on biosynthesis and secretion of nisin by *Lactococcus lactis*. *J. Biol. Chem.* **269**, 3555–3562 (1994).
- van der Meer, J. R. *et al.* Characterization of the *Lactococcus lactis* nisin A operon genes nisP, encoding a subtilisin-like serine protease involved in precursor processing, and nisR, encoding a regulatory protein involved in nisin biosynthesis. *J. Bacteriol.* **175**, 2578–2588 (1993).
- Newton, G. G., Abraham, E. P. & Berridge, N. J. Sulphur-containing amino-acids of nisin. *Nature* **171**, 606 (1953).
- Gross, E. & Morell, J. L. The number and nature of alpha, beta-unsaturated amino acids in nisin. *FEBS Lett* **2**, 61–64 (1968).
- Karakas Sen, A. *et al.* Post-translational modification of nisin. The involvement of NisB in the dehydration process. *Eur. J. Biochem.* **261**, 524–532 (1999).
- Garg, N., Salazar-Ocampo, L. M. & van der Donk, W. A. In vitro activity of the nisin dehydratase NisB. *Proc. Natl. Acad. Sci. U. S. A.* **110**, 7258–7263. <https://doi.org/10.1073/pnas.1222488110> (2013).
- Ortega, M. A. *et al.* Structure and mechanism of the tRNA-dependent lantibiotic dehydratase NisB. *Nature* **517**, 509–512. <https://doi.org/10.1038/nature13888> (2015).
- Koponen, O. *et al.* NisB is required for the dehydration and NisC for the lanthionine formation in the post-translational modification of nisin. *Microbiology* **148**, 3561–3568. <https://doi.org/10.1099/00221287-148-11-3561> (2002).
- Okeley, N. M., Paul, M., Stasser, J. P., Blackburn, N. & van der Donk, W. A. SpaC and NisC, the cyclases involved in subtilin and nisin biosynthesis, are zinc proteins. *Biochemistry* **42**, 13613–13624. <https://doi.org/10.1021/bi0354942> (2003).
- Li, B. & van der Donk, W. A. Identification of essential catalytic residues of the cyclase NisC involved in the biosynthesis of nisin. *J. Biol. Chem.* **282**, 21169–21175. <https://doi.org/10.1074/jbc.M701802200> (2007).
- Lubelski, J., Khusainov, R. & Kuipers, O. P. Directionality and coordination of dehydration and ring formation during biosynthesis of the lantibiotic nisin. *J. Biol. Chem.* **284**, 25962–25972. <https://doi.org/10.1074/jbc.M109.026690> (2009).
- Repka, L. M., Hetrick, K. J., Chee, S. H. & van der Donk, W. A. Characterization of leader peptide binding during catalysis by the nisin dehydratase NisB. *J. Am. Chem. Soc.* **140**, 4200–4203. <https://doi.org/10.1021/jacs.7b13506> (2018).
- Quiao, M. & Saris, P. E. Evidence for a role of NisT in transport of the lantibiotic nisin produced by *Lactococcus lactis* N8. *FEMS Microbiol. Lett.* **144**, 89–93 (1996).
- Higgins, C. F. ABC transporters: from microorganisms to man. *Annu. Rev. Cell Biol.* **8**, 67–113. <https://doi.org/10.1146/annurev.cb.08.110192.000435> (1992).
- Fath, M. J. & Kolter, R. ABC transporters: bacterial exporters. *Microbiol. Rev.* **57**, 995–1017 (1993).
- Hävarstein, L. S., Diep, D. B. & Nes, I. F. A family of bacteriocin ABC transporters carry out proteolytic processing of their substrates concomitant with export. *Mol. Microbiol.* **16**, 229–240 (1995).
- Schnell, N. *et al.* Analysis of genes involved in the biosynthesis of lantibiotic epidermin. *Eur. J. Biochem.* **204**, 57–68 (1992).
- Klein, C., Kaletta, C., Schnell, N. & Entian, K. D. Analysis of genes involved in biosynthesis of the lantibiotic subtilin. *Appl. Environ. Microbiol.* **58**, 132–142 (1992).
- Bierbaum, G., Brotz, H., Koller, K. P. & Sahl, H. G. Cloning, sequencing and production of the lantibiotic mersacidin. *FEMS Microbiol. Lett.* **127**, 121–126. <https://doi.org/10.1111/j.1574-6968.1995.tb07460.x> (1995).
- Siegers, K., Heinzmann, S. & Entian, K. D. Biosynthesis of lantibiotic nisin. Posttranslational modification of its prepeptide occurs at a multimeric membrane-associated lanthionine synthetase complex. *J. Biol. Chem.* **271**, 12294–12301 (1996).
- Kiesau, P. *et al.* Evidence for a multimeric subtilin synthetase complex. *J. Bacteriol.* **179**, 1475–1481 (1997).
- Nagao, J. *et al.* Localization and interaction of the biosynthetic proteins for the lantibiotic, Nukacin ISK-1. *Biosci. Biotechnol. Biochem.* **69**, 1341–1347. <https://doi.org/10.1271/bbb.69.1341> (2005).

31. Rink, R. *et al.* Lantibiotic structures as guidelines for the design of peptides that can be modified by lantibiotic enzymes. *Biochemistry* **44**, 8873–8882. <https://doi.org/10.1021/bi050081h> (2005).
32. Kluskens, L. D. *et al.* Post-translational modification of therapeutic peptides by NisB, the dehydratase of the lantibiotic nisin. *Biochemistry* **44**, 12827–12834. <https://doi.org/10.1021/bi050805p> (2005).
33. Rink, R. *et al.* NisC, the cyclase of the lantibiotic nisin, can catalyze cyclization of designed nonlantibiotic peptides. *Biochemistry* **46**, 13179–13189. <https://doi.org/10.1021/bi700106z> (2007).
34. Kuipers, A., Meijer-Wierenga, J., Rink, R., Kluskens, L. D. & Moll, G. N. Mechanistic dissection of the enzyme complexes involved in biosynthesis of lactacin 3147 and nisin. *Appl. Environ. Microbiol.* **74**, 6591–6597. <https://doi.org/10.1128/AEM.01334-08> (2008).
35. van den Berg van Saparoea, H. B., Bakkes, P. J., Moll, G. N. & Driessen, A. J. Distinct contributions of the nisin biosynthesis enzymes NisB and NisC and transporter NisT to prenisin production by *Lactococcus lactis*. *Appl. Environ. Microbiol.* **74**, 5541–5548. <https://doi.org/10.1128/AEM.00342-08> (2008).
36. Zhou, L., van Heel, A. J. & Kuipers, O. P. The length of a lantibiotic hinge region has profound influence on antimicrobial activity and host specificity. *Front. Microbiol.* **6**, 11. <https://doi.org/10.3389/fmicb.2015.00011> (2015).
37. van Heel, A. J. *et al.* Discovery, production and modification of five novel lantibiotics using the promiscuous nisin modification machinery. *ACS Synth. Biol.* **5**, 1146–1154. <https://doi.org/10.1021/acssynbio.6b00033> (2016).
38. Zhou, L., van Heel, A. J., Montalban-Lopez, M. & Kuipers, O. P. Potentiating the activity of nisin against *Escherichia coli*. *Front. Cell Dev. Biol.* **4**, 7. <https://doi.org/10.3389/fcell.2016.00007> (2016).
39. Lagedroste, M., Reiners, J., Smits, S. H. J. & Schmitt, L. Systematic characterization of position one variants within the lantibiotic nisin. *Sci. Rep.* **9**, 935. <https://doi.org/10.1038/s41598-018-37532-4> (2019).
40. Li, B. *et al.* Structure and mechanism of the lantibiotic cyclase involved in nisin biosynthesis. *Science* **311**, 1464–1467. <https://doi.org/10.1126/science.1121422> (2006).
41. Reiners, J., Abts, A., Clemens, R., Smits, S. H. & Schmitt, L. Stoichiometry and structure of a lantibiotic maturation complex. *Sci. Rep.* **7**, 42163. <https://doi.org/10.1038/srep42163> (2017).
42. Ra, S. R., Qiao, M., Immonen, T., Pujana, I. & Saris, E. J. Genes responsible for nisin synthesis, regulation and immunity form a regulon of two operons and are induced by nisin in *Lactococcus lactis* N8. *Microbiology* **142**(Pt 5), 1281–1288. <https://doi.org/10.1099/13500872-142-5-1281> (1996).
43. Izaguirre, G. & Hansen, J. N. Use of alkaline phosphatase as a reporter polypeptide to study the role of the subtilin leader segment and the SpaT transporter in the posttranslational modifications and secretion of subtilin in *Bacillus subtilis* 168. *Appl. Environ. Microbiol.* **63**, 3965–3971 (1997).
44. Meyer, C. *et al.* Nucleotide sequence of the lantibiotic Pep5 biosynthetic gene cluster and functional analysis of PepP and PepC. Evidence for a role of PepC in thioether formation. *Eur. J. Biochem.* **232**, 478–489 (1995).
45. Gross, E. & Morell, J. L. The presence of dehydroalanine in the antibiotic nisin and its relationship to activity. *J. Am. Chem. Soc.* **89**, 2791–2792 (1967).
46. Ingram, L. A ribosomal mechanism for synthesis of peptides related to nisin. *Biochim. Biophys. Acta* **224**, 263–265 (1970).
47. Khusainov, R., Heils, R., Lubelski, J., Moll, G. N. & Kuipers, O. P. Determining sites of interaction between prenisin and its modification enzymes NisB and NisC. *Mol. Microbiol.* **82**, 706–718. <https://doi.org/10.1111/j.1365-2958.2011.07846.x> (2011).
48. Khusainov, R. & Kuipers, O. P. The presence of modifiable residues in the core peptide part of precursor nisin is not crucial for precursor nisin interactions with NisB- and NisC. *PLoS ONE* **8**, e74890. <https://doi.org/10.1371/journal.pone.0074890> (2013).
49. Lenders, M. H., Beer, T., Smits, S. H. & Schmitt, L. In vivo quantification of the secretion rates of the hemolysin A Type I secretion system. *Sci. Rep.* **6**, 33275. <https://doi.org/10.1038/srep33275> (2016).
50. Zaitseva, J., Jenewein, S., Jumpertz, T., Holland, I. B. & Schmitt, L. H662 is the linchpin of ATP hydrolysis in the nucleotide-binding domain of the ABC transporter HlyB. *EMBO J* **24**, 1901–1910. <https://doi.org/10.1038/sj.emboj.7600657> (2005).
51. Choudhury, H. G. *et al.* Structure of an antibacterial peptide ATP-binding cassette transporter in a novel outward occluded state. *Proc. Natl. Acad. Sci. U. S. A.* **111**, 9145–9150. <https://doi.org/10.1073/pnas.1320506111> (2014).
52. Lin, D. Y., Huang, S. & Chen, J. Crystal structures of a polypeptide processing and secretion transporter. *Nature* **523**, 425–430. <https://doi.org/10.1038/nature14623> (2015).
53. Reimann, S. *et al.* Interdomain regulation of the ATPase activity of the ABC transporter haemolysin B from *Escherichia coli*. *Biochem. J.* **473**, 2471–2483. <https://doi.org/10.1042/BCJ20160154> (2016).
54. Bock, C., Zollmann, T., Lindt, K.-A., Tampé, R. & Abele, R. Peptide translocation by the lysosomal ABC transporter TAPL is regulated by coupling efficiency and activation energy. *Sci. Rep.* **9**, 11884. <https://doi.org/10.1038/s41598-019-48343-6> (2019).
55. Zheng, S., Nagao, J. I., Nishie, M., Zendo, T. & Sonomoto, K. ATPase activity regulation by leader peptide processing of ABC transporter maturation and secretion protein, NukT, for lantibiotic nukacin ISK-1. *Appl. Microbiol. Biotechnol.* <https://doi.org/10.1007/s00253-017-8645-2> (2017).
56. Morgan, J. L. W., Acheson, J. F. & Zimmer, J. Structure of a Type-1 secretion system ABC transporter. *Structure* **25**, 522–529. <https://doi.org/10.1016/j.str.2017.01.010> (2017).
57. Khusainov, R., Moll, G. N. & Kuipers, O. P. Identification of distinct nisin leader peptide regions that determine interactions with the modification enzymes NisB and NisC. *FEBS Open Bio* **3**, 237–242. <https://doi.org/10.1016/j.fob.2013.05.001> (2013).
58. Terzaghi, B. E. & Sandine, W. E. Improved medium for lactic streptococci and their bacteriophages. *Appl. Microbiol.* **29**, 807–813 (1975).
59. Jensen, P. R. & Hammer, K. Minimal requirements for exponential growth of *Lactococcus lactis*. *Appl. Environ. Microbiol.* **59**, 4363–4366 (1993).
60. Holo, H. & Nes, I. F. High-frequency transformation, by electroporation, of *Lactococcus lactis* subsp. *cremoris* grown with glycine in osmotically stabilized media. *Appl. Environ. Microbiol.* **55**, 3119–3123 (1989).
61. AlKhatib, Z. *et al.* Lantibiotic immunity: inhibition of nisin mediated pore formation by NisI. *PLoS ONE* **9**, e102246. <https://doi.org/10.1371/journal.pone.0102246> (2014).
62. AlKhatib, Z. *et al.* The C-terminus of nisin is important for the ABC transporter NisFEG to confer immunity in *Lactococcus lactis*. *Microbiol. Open* **3**, 752–763. <https://doi.org/10.1002/mbo3.205> (2014).
63. Infed, N., Hanekop, N., Driessen, A. J., Smits, S. H. & Schmitt, L. Influence of detergents on the activity of the ABC transporter LmrA. *Biochim. Biophys. Acta* **2313**–2321, 2011. <https://doi.org/10.1016/j.bbame.2011.05.016> (1808).

Acknowledgements

We thank Peter Tommes for support and analysis of MALDI-TOF-MS experiments. We are greatly obliged to Diana Kleinschrodt and Iris Fey (former Protein Production Facility of HH) for their support of cloning the constructs. We also thank Olivia Spitz, Ioannis Panetas and Didem Kaya for their support during the early stages of the project. We are indebted to all members of the Institute of Biochemistry for stimulating discussions and support during the project. This work was supported by the Deutsche Forschungsgemeinschaft (DFG Grant Schm1279/13-1 to L.S.).

Author contributions

M.L., S.H.J.S. and L.S. designed the experiments, M.L. performed the experiments, J.R. purified NisB and NisC, M.L., S.H.J.S. and L.S. analyzed and interpreted the data, M.L., S.H.J.S. and L.S. wrote and revised the manuscript.

Competing interests

The authors declare no competing interests.

Additional information

Supplementary information is available for this paper at <https://doi.org/10.1038/s41598-020-69225-2>.

Correspondence and requests for materials should be addressed to L.S.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2020